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Unmodified fluorescein as a fluorescent chemosensor for fluoride ion detection

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Abstract—Unmodified fluorescein (1) behaves as a fluorescent chemosensor for F^- detection, where the F^- -induced fluorescence enhancement is driven by a transfer of the phenolic OH protons to F^- . © 2007 Elsevier Ltd. All rights reserved.

The recognition and sensing of anions have attracted a great deal of attention because of biological and environmental importance of anions.¹ Fluoride ion (F^-) is one of the most important anions due to its pivotal roles in dental care and the treatment of osteoporosis.² Design of fluorescent probe for F^- detection has therefore attracted much attention due to high sensitivity and simplicity of the fluorescence analysis.³

Fluorescein is a dye used extensively as bio-labeling reagents and fluorescent probes due to its excellent photophysical properties, such as long-wavelength absorption and emission, high fluorescence quantum yield, and high stability against light.⁴ Considerable effort has been devoted to the development of fluorescent probes for reactive oxygen species $(ROS)^5$ and metal cations⁶ based on the fluorescein platform. There are, however, only three reports of fluorescein-based fluorescent anion sensor.⁷ Yoon et al.^{7a} synthesized a fluorescein derivative conjugated with boronic acid and aminomethyl groups. This material shows a F⁻induced emission enhancement due to a suppression of the photoinduced electron transfer from the amine nitrogen to the photoexcited fluorescein moiety by a cooperative coordination of F^- with the ligand groups. Yang et al.^{7b} synthesized a fluorescein derivative whose two phenolic OH are protected by tert-butyldimethylsilyl (TBS) groups. This shows a F⁻-induced fluorescence enhancement due to the deprotection of TBS, leading to

a formation of the spirocycle-opened 'emissive' fluorescein species. Very recently, a fluorescein bearing a thiourea group was synthesized by Kim et al.,^{7c} which shows an anion-induced fluorescence enhancement. In that, hydrogen-bonding interaction between anion and thiourea group leads to a spirocycle opening of the fluorescein moiety, resulting in fluorescence enhancement. Needless to say, practical anion sensing requires inexpensive and easily preparable sensors;⁸ however, all of these fluorescein-based anion sensors require a synthesis step for sensor preparation.⁷

Here we report that a commercially-available 'unmodified' fluorescein (1, Fig. 1) behaves as a fluorescent chemosensor for F^- detection, enabling selective $F^$ sensing among the halide anions. We describe here that the strong fluorescence enhancement of 1 by F^- is simply triggered by a transfer of the phenolic OH protons to F^- , leading to a formation of the spirocycle-opened emissive anionic species.

As shown in Figure 2, 1 (1 μ M) dissolved in acetonitrile (MeCN) is nonfluorescent. Addition of 10 equiv of F⁻ (as a *n*-Bu₄N⁺ salt) to the solution, however, leads to



Figure 1. Structures of fluorescein (1) and its derivatives (2, 3).

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Figure 2. Fluorescence spectra of 1 (1 μ M) in MeCN measured with 10 equiv of respective anions as *n*-Bu₄N⁺ salt ($\lambda_{ex} = 480$ nm). (Inset) change in fluorescence color.

an appearance of a strong green fluorescence at 500-600 nm. Moderate and weak emission enhancement is observed for AcO⁻ and H₂PO₄⁻, respectively, whereas no enhancement is observed for other anions $(Cl^{-}, Br^{-}, I^{-}, and HSO_{4}^{-})$. As shown in Figure 3, fluorescence titration with F^- reveals that the fluorescence enhancement is saturated upon the addition of 10 equiv of F^- , where the fluorescence quantum yield is 0.61.⁹ The emission enhancement is determined to be 2100fold, which is the highest value among the reported fluorescein-based fluorescent F⁻ sensors.⁷ In contrast, addition of even 100 equiv of other halide anions (Cl⁻, Br⁻, and I⁻) shows no emission enhancement (Fig. 3). Notably, as shown in Figure S1,¹⁰ the fluorescence response of 1 toward F^- is unaffected by the presence of other halide anions (Cl⁻, Br⁻, and I⁻), indicating that 1 is potentially available for selective F^- detection among the halide anions.

Figure 4 shows absorption spectra of 1 (10μ M) in MeCN. Without anions, 1 is colorless and exhibits almost no absorption at 400–550 nm, indicating that 1 exists as a spirocycle-closed form. This is confirmed by a distinctive spiro-carbon shift at 83.51 ppm in the ¹³C NMR spectrum of 1.¹¹ Addition of F⁻, however, leads



Figure 4. Absorption spectra of 1 (10 μ M) measured in MeCN with 4 equiv of respective anions. (Inset) Color change.

to an appearance of a strong absorption ($\lambda_{max} = 514 \text{ nm}$), along with a clear color change of the solution from colorless to yellow-green. Moderate and weak absorption increase is observed for AcO⁻ and H₂PO₄⁻, respectively, but almost no change is observed for other anions (Cl⁻, Br⁻, I⁻, and HSO₄⁻).

As shown in Figure 5, absorption titration of 1 with F^- reveals that addition of first 2 equiv of F^- leads to an increase in the entire absorption at 400–550 nm (see blue line). Further addition of 2 equiv of F^- (see red line) leads to further increase in 465–550 nm absorbance, but also leads to a decrease in 400–465 nm absorbance with an isosbestic point at 465 nm. These findings imply that two kinds of fluorescein species form in response to the interaction between 1 and F^- with 1:2 and 1:4 stoichiometry. This is confirmed by the Job's plots of 1 with F^- : as shown in Figure 6, 460 nm absorbance shows a maximum at X (=[F^-]/([F^-] + [1])) = 0.66, whereas



Figure 3. Fluorescence titration of **1** (1 μ M) with F⁻ ($\lambda_{ex} = 480$ nm). (Inset) Change in fluorescence intensity ($\lambda_{em} = 532$ nm).



Figure 5. Absorption titration of 1 (10 μ M) with F⁻. (Inset) Change in absorbance monitored at 514 nm. The detailed absorption spectrum change: see Figure S2.¹⁰



Figure 6. Job's plot for F^- versus 1 in MeCN measured (closed circle) at 514 nm and (open circle) at 460 nm. $[F^-] + [1] = 20 \ \mu M$.

514 nm absorbance shows a maximum at X = 0.8. The respective absorption spectra of 1 obtained with 2 and 4 equiv of F⁻ (Fig. 5) are similar to those of the monoanion and dianion species of 1 observed in water.¹² As reported, ^{12a,13} 1 is emissive in the anionic forms, especially in the dianion form. These findings suggest that, as proposed in Scheme 1, two phenolic OH protons of 1 are removed via the interaction with F⁻ in the ground state, resulting in the formation of 'emissive' two anionic species.

For further confirmation of the proposed F^- sensing mechanism of 1, ¹H NMR titration of 1 was carried out with F^- in DMSO- d_6 (Figure S3¹⁰). Upon the addition of 0.2 equiv of F^- , phenolic OH proton of 1 (10.09 ppm) shifts downfield (11.02 ppm) with a significant intensity decrease. This indicates the occurrence of a hydrogen-bonding interaction between the phenolic OH protons of 1 and F⁻.¹⁴ Addition of 2 equiv of F⁻ leads to a complete disappearance of the OH proton. In addition, all of the aromatic protons of **1** shift upfield. This means that net electron density of the aromatic ring of 1 increases,¹⁵ indicative of the removal of phenolic OH proton from 1. The upfield shift of the aromatic protons almost stops upon the addition of 4 equiv of F^- , meaning that the two phenolic OH protons of 1 are completely removed at this stage. This result is consistent with the absorption titration and the Job's plot data (Figs. 5 and 6). In addition, upon the addition of >4 equiv of F⁻, a new triplet signal appears at 16.1 ppm (J = 121 Hz), which is ascribed to a FHF⁻ dimer,^{1h,3d,g,16} meaning that the phenolic OH protons of 1 are actually removed by F⁻. The removal of the phenolic OH protons is also confirmed by ESI-MS analysis (Figure $\hat{S}4^{10}$): the MS chart obtained with 1 and 5 equiv of F⁻ in MeCN shows a strong peak at m/z1056.8, which is ascribed to $([1-2H] + 3[n-Bu_4N])^{\dagger}$



Scheme 1. Proposed F⁻ sensing mechanism of 1.

ion, in which both phenolic OH protons of **1** are removed. This means that anionic species of **1** are actually produced by the F^- -induced proton removal. The F^- sensing mechanism of **1** can therefore be summarized as Scheme 1: first 2 equiv of F^- leads to removal of one phenolic OH proton of **1** via a hydrogen-bonding interaction, resulting in a formation of a FHF⁻ dimer and the emissive monoanion of **1**.¹⁵ Another phenolic OH proton of **1** equiv of F^- , forming FHF⁻ dimer and the emissive dianion.

The proposed F^- sensing mechanism of 1 (Scheme 1) is further confirmed by absorption and fluorescence behaviors of the control compounds, 2 and 3 (Fig. 1; see Materials¹⁰). Compound 2 has a spirocycle-open form by the esterification of the carboxylic acid group of 1; therefore, even without anions, 2 exhibits an absorption at 400–500 nm (Figure S5¹⁰). This absorption spectrum is similar to that of 1 obtained with <2 equiv of F⁻ (Fig. 5), meaning that the removal of the phenolic OH proton of 1 actually leads to the spirocycle opening (Scheme 1). Upon the addition of F^- to 2, a new absorption appears at 524 nm, along with a decrease in 400-475 nm absorbance (isosbestic point: 475 nm), as is also the case for 1 with 2–4 equiv of F^{-} . This means that the phenolic OH proton of 2 is also removed by F^- . Job's plot of **2** with F^- (Figure S6¹⁰) shows a maximum absorption at X = 0.66, indicating that the proton removal of 2 by F^- occurs in a 1:2 stoichiometry.¹⁷ These indicate that 2 equiv of F⁻ leads to removal of one phenolic OH proton; this fact supports the proposed deprotonation mechanism of 1 (Scheme 1). Compound 2 is weakly fluorescent, but addition of F⁻ leads to a fluorescence enhancement (Figure S7¹⁰).¹⁸ This means that the removal of the phenolic OH proton produces stronger emitting species; this also supports the proposed mechanism (Scheme 1).

Compound 3 (Fig. 1) has a spirocycle-open form, where both carboxylic acid and phenolic OH groups of 1 are methylated. This compound does not show any absorption or fluorescence response to F^- (Figure S10¹⁰). This is because 3 does not produce anionic species due to the lack of a phenolic OH proton. This finding again support the proposed mechanism: the removal of the phenolic OH proton of 1 by F^- triggers the strong fluorescence enhancement (Scheme 1).

In conclusion, we found that the unmodified 'readymade' fluorescein (1) behaves as a fluorescent F^- sensor, which shows potential for selective F^- detection among the halide anions. As is usually observed for the fluorescent anion sensors,^{3,7} the present fluorescein system has difficulty in the application for aqueous samples.¹⁹ However, the fluorescein is photoexcited by visible light and shows high fluorescence quantum yield and high sensitivity; therefore, the fluorescein may be applicable as a fluorescent F^- sensor. The sensing mechanism clarified here, which cleverly detects F^- by simple proton transfer processes, may contribute to the design of more effective and more sensitive fluorescent anion sensor based on the fluorescein platform.

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Supplementary data

Supplementary data (Materials and Figures S1–S16). Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet. 2007.10.086.

References and notes

- (a) Schmidtchen, F. P.; Berger, M. Chem. Rev. 1997, 97, 1609–1646; (b) Snowden, T. S.; Anslyn, E. V. Curr. Opin. Chem. Biol. 1999, 3, 740–746; (c) Beer, P. D.; Gale, P. A. Angew. Chem., Int. Ed. 2001, 40, 486–516; (d) Sessler, J. L.; Davis, J. M. Acc. Chem. Res. 2001, 34, 989–997; (e) Gale, P. A. Coord. Chem. Rev. 2001, 213, 79–128; (f) Martínez-Máñez, R.; Sancanón, F. Chem. Rev. 2003, 103, 4419–4476; (g) Amendola, V.; Bonizzoni, M.; Esteban-Gómez, D.; Fabbrizzi, L.; Licchelli, M.; Sancenón, F.; Taglietti, A. Coord. Chem. Rev. 2006, 250, 1451–1470; (h) Gunnlaugsson, T.; Glynn, M.; Tocci, G. M.; Kruger, P. E.; Pfeffer, F. M. Coord. Chem. Rev. 2006, 250, 3094–3117.
- (a) Kirk, K. L. Biochemistry of the Halogens and Inorganic Halides; Plenum: New York, 1991, p 58; (b) Kleerekoper, M. Endocrinol. Metab. Clin. North Am. 1998, 27, 441–452.
- 3. (a) de Silva, A. P.; Gunaratne, H. Q. N.; Gunnlaugsson, T.; Huxley, A. J. M.; McCoy, C. P.; Rademacher, J. T.; Rice, T. E. *Chem. Rev.* **1997**, *97*, 1515–1566; (b) Cho, E. J.; Moon, J. W.; Ko, S. W.; Lee, J. Y.; Kim, S. K.; Yoon, J.; Nam, K. C. J. Am. Chem. Soc. 2003, 125, 12376-12377; (c) Xu, G. X.; Tarr, M. A. Chem. Commun. 2004, 1050-1051; (d) Peng, X.-J.; Wu, Y.-K.; Fan, J.-L.; Tian, M.-Z.; Han, K.-L. J. Org. Chem. 2005, 70, 10524-10531; (e) Lin, Z.-H.; Ou, S.-J.; Duan, C.-Y.; Zhang, B.-G.; Bai, Z.-P. Chem. Commun. 2006, 624-626; (f) Liu, X. Y.; Bai, D. R.; Wang, S. Angew. Chem., Int. Ed. 2006, 45, 5475-5478; (g) Lin, C.-I.; Selvi, S.; Fang, J.-M.; Chou, P.-T.; Lai, C.-H.; Cheng, Y.-M. J. Org. Chem. 2007, 72, 3537-3542; (h) Kim, S. Y.; Hong, J.-I. Org. Lett. 2007, 9, 3109-3112; (i) Jiang, X.; Vieweger, M. C.; Bollinger, J. C.; Dragnea, B.; Lee, D. Org. Lett. 2007, 9, 3579-3582; (j) Hirano, J.; Miyata, H.; Hamase, K.; Zaitsu, K. Tetrahedron Lett. 2007, 48, 4861-4864.
- 4. Lakowicz, J. R. *Principles of Fluorescence Spectroscopy*, 3rd ed.; Springer: New York, 2006; pp 67–69.
- (a) Chang, M. C. Y.; Pralle, A.; Isacoff, E. Y.; Chang, C. J. J. Am. Chem. Soc. 2004, 126, 15392–15393; (b) Maeda, H.; Yamamoto, K.; Kohno, I.; Hafsi, L.; Itoh, N.; Nakagawa, S.; Kanagawa, N.; Suzuki, K.; Uno, T. Chem. Eur. J. 2007, 13, 1946–1954.
- (a) Minta, A.; Kao, J. P. Y.; Tsien, R. Y. J. Biol. Chem. 1989, 264, 8171–8178; (b) Walkup, G. K.; Burdette, S. C.; Lippard, S. J.; Tsien, R. Y. J. Am. Chem. Soc. 2000, 122,

5644–5645; (c) Hirano, T.; Kikuchi, K.; Urano, Y.; Higuchi, T.; Nagano, T. J. Am. Chem. Soc. 2000, 122, 12399–12400; (d) Komatsu, H.; Iwasawa, N.; Citterio, D.; Suzuki, Y.; Kubota, T.; Tokuno, K.; Kitamura, Y.; Oka, K.; Suzuki, K. J. Am. Chem. Soc. 2004, 126, 16353– 16360; (e) Nolan, E. M.; Jaworski, J.; Racine, M. E.; Sheng, M.; Lippard, S. J. Inorg. Chem. 2006, 45, 9748– 9757.

- (a) Swamy, K. M. K.; Lee, Y. J.; Lee, H. N.; Chun, J.; Kim, Y.; Kim, S.-J.; Yoon, J. J. Org. Chem. 2006, 71, 8626–8628; (b) Yang, X.-F.; Ye, S.-J.; Bai, Q.; Wang, X.-Q. J. Fluoresc. 2007, 17, 81–87; (c) Wu, J.-S.; Kim, H. J.; Lee, M. H.; Yoon, J. H.; Lee, J. H.; Kim, J. S. Tetrahedron Lett. 2007, 48, 3159–3162.
- Miyaji, H.; Sessler, J. L. Angew. Chem., Int. Ed. 2001, 40, 154–157.
- 9. The fluorescence quantum yield of fluorescein (0.85 in 0.1 M NaOH aqueous solution) was used as a standard: Parker, C. A.; Rees, W. T. *Analyst* **1960**, *85*, 587–600, The measurement was carried out at 300 K in an aerated condition. The fluorescence quantum yield of fluorescein is insensitive to oxygen.
- 10. See Supplementary data.
- DBS Web: http://www.aist.go.jp/RIODB/SDBS/, SDBS No: 6347, National Institute of Advanced Industrial Science and Technology (AIST), Japan.
- (a) Klonis, N.; Sawyer, W. H. J. Fluoresc. 1996, 6, 147– 157; (b) Mchedlov-Petrossyan, N. O.; Mayorga, R. S. J. Chem. Soc., Faraday Trans. 1992, 88, 3025–3032; (c) Mota, M. C.; Carvalho, P.; Ramalho, J.; Leite, E. Int. Ophthalmol. 1991, 15, 321–326.
- (a) Urano, Y.; Kamiya, M.; Kanda, K.; Ueno, T.; Hirose, K.; Nagano, T. J. Am. Chem. Soc. 2005, 127, 4888–4894;
 (b) Ghislain, G.; Rene, A.; Jacques, L. J. Chim. Phys. 1975, 72, 647–653.
- Zhang, X.; Guo, L.; Wu, F.-Y.; Jiang, Y.-B. Org. Lett. 2003, 5, 2667–2670.
- (a) Boiocchi, M.; Del Boca, L.; Gomez, D. E.; Fabbrizzi, L.; Licchelli, M.; Monzani, E. J. Am. Chem. Soc. 2004, 126, 16507–16514; (b) Amendola, V.; Gómez, D. E.; Fabbrizzi, L.; Licchelli, M. Acc. Chem. Res. 2006, 39, 343– 353.
- Descalzo, A. B.; Rurack, K.; Weisshoff, H.; Martínez-Máñez, R.; Marcos, M. D.; Amorós, P.; Hoffmann, K.; Soto, J. J. Am. Chem. Soc. 2005, 127, 184–200.
- 17. The formation of the anion form of compound **2** is confirmed by ¹H NMR titration (Fig. S8) and ESI-MS (Fig. S9) analyses. Upon the addition of F⁻, the phenolic OH proton of **2** at 11.06 ppm (in DMSO-*d*₆) disappears and all of the aromatic protons shift upfield, as is also the case for **1**. Triplet FHF⁻ dimer signals also appear at 16.1 ppm (J = 121 Hz). ESI-MS analysis of a MeCN solution containing **2** with 5 equiv of F⁻ shows a peak at m/z 843.5, assigned to ($[2-1H] + 2[n-Bu_4N]$)⁺ ion. These data clearly support the formation of the anion form of **2** by the F⁻-induced removal of the phenolic OH proton.
- 18. The fluorescence quantum yield of $\mathbf{2}$ with 5 equiv of F^- is only 0.14.
- 19. Addition of 0.1% or 1% water to MeCN containing fluorescein and 20 equiv of F⁻ shows fluorescence intensity similar to that obtained without water, although the addition of 5% water to MeCN shows almost zero fluorescence intensity.